

# Stool Culture, Collection Instructions

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## Specimen Collection - Enteric Transport Container (**Green Cap / Red Fluid**):

The patient must not be taking any form of medication unless specifically permitted by the physician. The use of barium and bismuth with certain radiological examinations is particularly damaging to the specimen. Collection should be at least one week after usage.

Samples from multiple stool specimens are recommended to assure optimum recovery of enteric pathogens. **Three consecutive specimens** should be obtained during the acute stage. To identify chronic, subclinical, and carrier states, collection of stool specimens on three consecutive days is recommended.

1. The bowel movement should be passed into a clean, dry, wide-mouthed container. Urine or water must not contaminate the stool.
2. Select watery, bloody, or mucoid portions of the stool using the attached collection spoon. To insure adequate sampling of formed stool, sample portions of the side, middle, and end of the bolus.
3. Fill two enteric transport vials. Place enough specimen into the vial to raise the level of the medium to the **FILL** line on the vial.
4. Mash and mix the stool against the side of the vial with the spoon. Tighten the cap and shake the vial until the mixture appears homogeneous.
5. Two enteric transport vials are needed for the stool culture.
  - a. Label the vials with pertinent patient information. Include the time and date of collection. Label one green top with "EHEC" and refrigerate.
  - b. Label the second green top with "Stool Culture" and keep at room temperature.
6. Return Specimens to a Weland Clinical Laboratories site within **48 hours** of collection.